Opinion

Nutritional Immunology: A Multi-Dimensional Approach

Fleur Ponton^{1*}, Kenneth Wilson², Sheena C. Cotter³, David Raubenheimer⁴, Stephen J. Simpson¹

1 School of Biological Sciences, The University of Sydney, Sydney, New South Wales, Australia, 2 Lancaster Environment Centre, Lancaster University, Lancaster, United Kingdom, 3 School of Biological Sciences, Queen's University, Belfast, United Kingdom, 4 Institute of Natural Sciences, Massey University, Auckland, New Zealand

Nutrition is critical to immune defence and resistance to pathogens, with consequences that affect the health, welfare, and reproductive success of individual organisms [1,2], and also has profound ecological and evolutionary implications [3-5]. In humans, under-nutrition, notably of protein, is a major contributor to morbidity and mortality due to infectious diseases, particularly in the developing world [1]. Likewise, over-nutrition and its associated metabolic disorders may impair immune function, disrupt the relationship with symbiotic and commensal microbiota, and increase susceptibility to infectious disease [6]. Despite the undoubted importance of nutrition to immune defence, the challenge remains to capture the complexity of this relationship. There are three main aspects to this complexity: (i) nutrition is a complex multi-dimensional problem for hosts, pathogens, and commensals; (ii) host immunity is a complex, multi-dimensional trait; and (iii) nutrition and immunity interact via multiple direct and indirect pathways, including involvement of the host's microbiota.

Nutrition Is a Multi-Dimensional Problem for Hosts, Pathogens, and Commensal Organisms

Although widely used, the terms "overnutrition" and "under-nutrition" are rarely defined in studies, and often the key nutritional variables have not been identified. Many studies consider foods as uniform commodities and manipulate the amount available without considering the food's nutritional composition or having a quantitative understanding of the animal's nutrient requirements (see for instance [7,8]). Other studies focus on a single dietary attribute (typically its calorie content) or nutritional component (e.g., the amount of protein or nitrogen in the diet), and experimentally manipulate this whilst maintaining other dietary components at a constant level, thus confounding changes in the focal nutrient/attribute with changes in the ratio of nutrients in the diet [9-11]. Although these studies have had their successes, we believe that this single currency approach provides only a limited understanding of the nutritional constraints on pathogen defence (see also [2]). The Geometric Framework (GF) [12-14] was specifically developed to capture these multi-dimensional aspects of nutrition and offers promise for the study of nutritional immunology, allowing quantitative predictions that can be statistically tested. The GF identifies nutritional optima (intake and growth targets) in multi-dimensional nutritional space and thus provides a rigorous definition and quantification of "under"- and "over"nutrition, as well as a mean of associating immune responses, host performance, responses of host microbial communities, and pathogen growth rates with particular nutritional states.

The importance of considering the simultaneous and interactive effects of multiple nutrients when studying immune function has been illustrated by a number of studies on insects and rodents [15–18]. For example, Peck et al. [18] found that mice survived better on diets containing a higher ratio of protein (P) to carbohydrate (C) following inoculation with *Salmonella typhimurium*. Similarly, the ability of caterpillars to resist viral and bacterial infection increased as dietary P:C rose, and infected insects selected a higher protein diet, indicating a form of nutritional self-medication [15–17].

Hosts are not the only organisms facing the complexity of nutrition. Parasites and pathogens rely on the host for provision of resources and may not share the same nutritional requirements, setting up the potential for resource competition and manipulation between the different parties [18–20]. The complexity of the nutritional interactions between hosts and pathogens is made substantially greater by the fact that animals play host not only to invading pathogens, but also to entire communities of commensal and symbiotic microorganisms that receive their nutrition from the host and in turn contribute essential nutrients and play a role in immune defence [21-24]. Gut microbiota have been shown to have profound and unanticipated effects on immune defence and inflammatory responses [23,25-27], and in mammals, disturbances of the gut microbiota have been implicated in diseases such as obesity, type 1 diabetes, and various cancers [21,23,28]. Furthermore, diet has a strong effect on the gut microbiota [29-32], both by serving as a vector for microorganisms and by affecting the physical, chemical, and structural properties of the gut [33-37].

The Immune System Has Multiple Components That React Differently to Nutrients

Immune loci are the most gene-dense regions of the genome in vertebrates (e.g., [38]), and even in insects, which lack an adaptive immune response and rely solely on the innate immune system, there are abundant components to the immune response, each designed to meet particular types of immune challenge [39]. It has recently been discovered using GF designs that immune components respond differently to host nutritional state. Cotter et al. [40] restricted control and immune-challenged caterpillars to one of 20 diets

Citation: Ponton F, Wilson K, Cotter SC, Raubenheimer D, Simpson SJ (2011) Nutritional Immunology: A Multi-Dimensional Approach. PLoS Pathog 7(12): e1002223. doi:10.1371/journal.ppat.1002223

Editor: Glenn F. Rall, The Fox Chase Cancer Center, United States of America

Published December 1, 2011

Copyright: © 2011 Ponton et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: FP was supported by a postdoctoral grant from The University of Sydney. SJS was supported by ARC federation and Laureate Fellowships, Australia. SCC was an NERC fellow. KW was funded by a Royal Society Travel For Collaboration grant and a University of Sydney Visiting Research Fellowship. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: fleur_ponton@hotmail.com

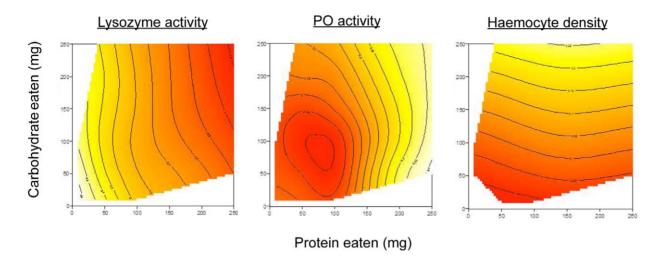


Figure1. Response landscapes for three immune traits in caterpillars fed one of 20 diets differing in the ratios and amounts of protein and carbohydrate. Adapted after Cotter et al. [40]. doi:10.1371/journal.ppat.1002223.g001

varying in both the quantity and ratio of P and C. Statistical analysis of response surfaces (Figure 1) showed that immune traits are differentially affected by macronutrient intake and that no diet can simultaneously optimize all components of the immune system (see below). Variation in these different traits has been shown to be repeatable and heritable [41,42] and linked to functional immune outcomes [43,44]. This raises the intriguing prospect that an animal might adjust its food selection to support immune components that best resist a given infection and perhaps also support a healthy microbial community.

A Framework to Address the Complexity of Nutritional Immunology

When the above mentioned complexities are considered, it becomes clear that an understanding of nutritional immunology must take account of a web of

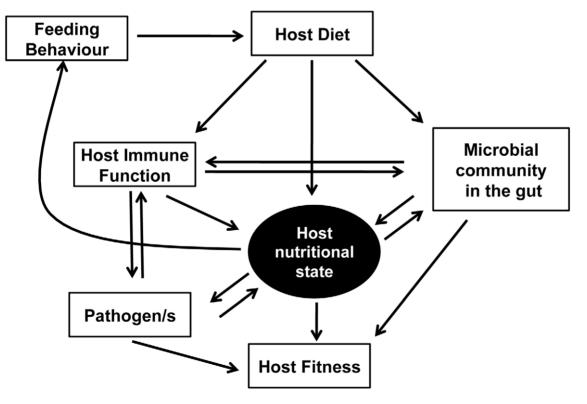


Figure 2. The network of interactions between nutrition and immunity. Diet affects host nutritional state and immune status, both of which interact with microbial symbionts, commensals, and pathogens to affect the fitness of all partners. Because nutrient feedbacks modulate host feeding behaviour, the potential exists for the host to adjust its diet to optimise its microbial interactions and increase resistance to infection. Alternatively, parasites and pathogens might subvert host feeding behaviour to their nutritional advantage. doi:10.1371/journal.ppat.1002223.g002

interactions between components. These include the nutritional quality of the diet, host feeding behaviour, host nutritional state, the growth of pathogen populations, the host-associated microbial community, multiple measures of host immune function and, ultimately, evolutionary considerations such as host and microbial fitness and selection processes (Figure 2).

When exploring this network of interactions, the first step (primary manipulation) is to define the effects of nutrition on the network. The recent study of Lee et al. [45] on Drosophila offers an example of how GF designs might be used in such an analysis of nutritional immunology. In that study, systematically varying the protein and carbohydrate content allowed variables including host lifespan and lifetime egg production to be mapped as response surfaces onto nutrient intake arrays, thereby parsing the consequences of nutritional state on these key life-history traits and providing a baseline for detailed physiological and molecular analysis. Using the same technique, Cotter et al. [40] (see

References

- Cunningham-Rundles S, McNeeley DF, Moon A (2005) Mechanisms of nutrient modulation of the immune response. J Allergy Clin Immunol 115: 1119–1128.
- Schneider D (2009) Physiological integration of innate immunity. In: Rolff J, Reynolds SE, eds. Insect infection and immunity. Oxford: Oxford University Press. pp 106–116.
- Sheldon BC, Verhulst S (1996) Ecological immunology: costly parasite defences and trade-offs in evolutionary ecology. Trends Ecol Evol 11: 317–321.
- Rolff J, Siva-Jothy MT (2003) Invertebrate ecological immunology. Science 301: 472–475.
- Hawley DM, Altizer SM (2011) Disease ecology meets ecological immunology: understanding the links between organismal immunity and infection dynamics in natural populations. Func Ecol 25: 48–60.
- Amar S, Zhou Q, Shaik-Dasthagirisaheb Y, Leeman S (2007) Diet-induced obesity in mice causes changes in immune responses and bone loss manifested by bacterial challenge. Proc Natl Acad Sc U S A 104: 20466–20471.
- Moret Y, Schmid-Hempel P (2000) Survival for immunity: the price of immune system activation for bumblebee workers. Science 290: 1166– 1168.
- Siva-Jothy MT, Thompson JJW (2002) Shortterm nutrient deprivation affects immune function. Physiol Entomol 27: 206–212.
- Popham HJR, Shelby KS (2009) Ascorbic acid influences the development and immunocompetence of larval *Heliothis virescens*. Entomol Exp Appl 133: 57–64.
- Popham HJR, Shelby KS (2007) Effect of inorganic and organic forms of selenium supplementation on development of larval *Heliothis* virescens. Entomol Exp Appl 125: 171–178.
- Popham HJR, Shelby KS, Popham TW (2005) Effect of dietary selenium supplementation on resistance to baculovirus infection. Biol Control 32: 419–426.
- Raubenheimer D, Simpson SJ (1993) The geometry of compensatory feeding in the locust. Anim Behav 45: 953–964.

above) mapped several immune traits onto P-C intake arrays (Figure 1), providing evidence that immune components responded in a nutrient-specific manner. Whether these different responses were driven by differing nutritional demands of the various immune traits, direct effects of nutrition on patterns of immune gene expression, or an indirect effect of changes to microbial communities in the gut or elsewhere in the body, remains to be discovered.

A more complete study of nutritional immunology would require including response surfaces for gut and body microbial communities, as well as a more detailed assessment of immune pathways, e.g., IMD and Toll antimicrobial peptide pathways. Having quantified the effects of diet composition, the responses of the network to perturbations could then be measured. This could be done by inoculating hosts with pathogens that challenge different components of the innate immune system; by using host strains deficient in different components of the

- Simpson SJ, Raubenheimer D (1993) A multilevel analysis of feeding-behavior - the geometry of nutritional decisions. Philos Trans R Soc B 342: 381–402.
- Simpson SJ, Raubenheimer D (2012) The nature of nutrition: a unifying framework from animal adaptation to human obesity. Princeton: Princeton University Press. In press.
- Lee KP, Cory JS, Wilson K, Raubenheimer D, Simpson SJ (2006) Flexible diet choice offsets protein costs of pathogen resistance in a caterpillar. Proc Biol Sci 273: 823–829.
- Povey S, Cotter SC, Simpson SJ, Lee KP, Wilson K (2009) Can the protein costs of bacterial resistance be offset by altered feeding behaviour? J Anim Ecol 78: 437–446.
- Raubenheimer D, Simpson SJ (2009) Nutritional PharmEcology: doses, nutrients, toxins, and medicines. Integr Comp Biol 49: 329–337.
- Peck MD, Babcock GF, Alexander JW (1992) The role of protein and calorie restriction in outcome from *Salmonella* infection in mice. J Parenter Enteral Nutr 16: 561–565.
- Smith VH, Holt RD (1996) Resource competition and within-host disease dynamics. Trends Ecol Evol 11: 386–389.
- Ponton F, Lalubin F, Fromont C, Wilson K, Behm C, et al. (2011) Hosts use altered macronutrient intake to circumvent parasite-induced reduction in fecundity. Int J Parasitol 41: 43– 50.
- Topping DL, Clifton PM (2001) Short-chain fatty acids and human colonic function: roles of resistant starch and nonstarch polysaccharides. Physiol Rev 81: 1031–1064.
- 22. Douglas AE (2010) The symbiotic habit. Princeton: Princeton University Press.
- Wen L, Ley RE, Volchkov PY, Stranges PB, Avanesyan L, et al. (2008) Innate immunity and intestinal microbiota in the development of Type 1 diabetes. Nature 455: 1109–1113.
- Kau AL, Ahern PP, Griffin NW, Goodman AL, Gordon JI (2011) Human nutrition, the gut microbiome and the immune system. Nature 474: 327–36.

immune response; using RNAi to knockdown particular immune genes; or by manipulating the commensal microbiota through antibiotic treatment. Finally, hosts could be offered the opportunity to express nutritional self-medication in experimental designs in which they are offered a choice of nutritionally complementary foods [45]. Studying the individual components of this complex interaction will allow us to formulate null models against which specific hypotheses can be formulated and tested.

Considering the complex nature of nutritional immunology, we argue that a description of the network of interactions that define the relationships between nutrition, immune function, infection, and microbiota is essential to provide a more comprehensive and robust understanding of the key determinants of the outcome of host-pathogen interactions. The GF provides a powerful organising framework for achieving such a synthesis.

- Salzman NH (2011) Microbiota-immune system interaction: an uneasy alliance. Curr Opin Microbiol 14: 99–105.
- Mazmanian SK, Round JL, Kasper DL (2008) A microbial symbiosis factor prevents intestinal inflammatory disease. Nature 453: 620–625.
- Ryu J-H, Ha E-M, Lee W-J (2010) Innate immunity and gut-microbe mutualism in *Drosophila*. Dev Comp Immunol 34: 369–376.
- Sekirov I, Russell SL, Antunes LCM, Finlay BB (2010) Gut microbiota in health and disease. Physiol Rev 90: 859–904.
- 29. De Filippo C, Cavalieri D, Di Paola M, Ramazzotti M, Poullet JB, et al. (2010) Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. Proc Natl Acad Sci U S A 107: 14691–14696.
- Turnbaugh PJ, Ridaura VK, Faith JJ, Rey FE, Knight R, et al. (2009) The effect of diet on the human gut microbiome: a metagenomic analysis in humanized gnotobiotic mice. Sci Transl Med 1: 6ra14.
- Ley RE, Hamady M, Lozupone C, Turnbaugh PJ, Ramey RR, et al. (2008) Evolution of mammals and their gut microbes. Science 320: 1647–1651.
- Muegge BD, Kuczynski J, Knights D, Clemente JC, González A, et al. (2011) Diet drives convergence in gut microbiome functions across mammalian phylogeny and within humans. Science 332: 970–974.
- Sørensen A, Mayntz D, Simpson SJ, Raubenheimer D (2010) Dietary ratio of protein to carbohydrate induces plastic responses in the gastrointestinal tract of mice. J Comp Physiol B 180: 259–266.
- Flint HJ, Duncan SH, Scott KP, Louis P (2007) Interactions and competition within the microbial community of the human colon: links between diet and health. Environ Microbiol 9: 1101–1111.
- Duncan SH, Lobley GE, Holtrop G, Ince J, Johnstone AM, et al. (2008) Human colonic microbiota associated with diet, obesity and weight loss. Int J Obes 32: 1720–1724.
- 36. Clissold FJ, Tedder BJ, Conigrave AD, Simpson SJ (2010) The gastrointestinal tract as

a nutrient-balancing organ. Proc R Soc B 1688: 1751–1759.

- Raubenheimer D, Bassil K (2007) Separate effects of macronutrient concentration and balance on plastic gut responses in locusts. J Comp Physiol B 177: 849–855.
- Beck S, Geraghty D, Inoko H, Rowen L (1999) Complete sequence and gene map of a human major histocompatibility complex. Nature 401: 921–923.
- Siva-Jothy MT, Moret Y, Rolff J (2005) Insect immunity: An evolutionary ecology perspective. Adv Insect Physiol 32: 1–48.
- Cotter SC, Simpson SJ, Raubenheimer D, Wilson K (2010) Macronutrient balance mediates trade-offs between immune function and life history traits. Func Ecol 25: 186–198.
- Cotter SC, Kruuk LEB, Wilson K (2004) Costs of resistance: genetic correlations and potential trade-offs in an insect immune System. J Evol Biol 17: 421–429.
- Cotter SC, Hails RS, Cory JS, Wilson K (2004) Density-dependent prophylaxis and conditiondependent immune function in Lepidopteran larvae: a multivariate approach. J Anim Ecol 73: 283–293.
- Wilson K, Cotter SC, Reeson AF, Pell JK (2001) Melanism and disease resistance in insects. Ecol Lett 4: 637–649.
- Wilson K (2005) Evolutionary ecology of insect host-parasite interactions: an ecological immunology perspective. Wallingford: CABI Publishing. pp. pp 289–246.
- Lee KP, Simpson SJ, Clissold FJ, Brooks R, Ballard JW, et al. (2008) Lifespan and reproduction in *Drosophila*: New insights from nutritional geometry. Proc Natl Acad Sci U S A 105: 2498–2503.