Supporting Online Material for

- 2 Wolbachia in a Major African Crop Pest Increases Susceptibility to Viral
- 3 Disease Rather than Protects
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Table S1. Determinants of overt viral deaths in *S. exempta* across outbreaks. 24

Term	Coef	s.e.m.	d.f.	Δ deviance	F	<i>P</i> -value
Season *	-2.799	0.272	1	127.27	13.01	0.0007
Head-capsule width (mean)	4.717	0.558	1	139.18	14.23	0.0004
Wolbachia prevalence †	4.195	0.436	1	117.44	12.01	0.0011
Excluded terms						
Larval density			1	24.63	3.49	0.0680
Outbreak number			1	6.52	0.62	0.4363
Date ‡			1	11.56	1.17	0.2847

^{*} model simplification revealed two levels: 2007/08 vs 2009/10 and 2010/11; † square-root 25 26

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The dependent variable in this analysis was the prevalence of virus-dead larvae observed across each of 59 outbreaks, where a minimum of thirty standardized counts were conducted; eight observations were excluded due to missing data. The proportion of dead larvae across all counts was weighted by the total number of larvae counted to control for variation in sampling intensity. Analysis was by generalized linear model with binomial errors and logit link function. To account for over-dispersion, an empirical dispersion parameter and F-tests were used. Parameter estimates in the minimal model are for explanatory terms when fitted last in the model. All other terms were excluded with P > 0.05 when fitted last in the model. Final model: intercept = -15.918 ± 1.244 ; null deviance = 848.60 (50 d.f.); residual deviance = 312.62 (47 d.f.)d.f.). None of the interactions between the main effects were statistically significant.

transformed; ‡ days since January 1st of calendar year.

Table S2. Determinants of overt viral deaths in *S. exempta* across outbreaks.

Term	Coef	s.e.m.	d.f.	Δ deviance	F	<i>P</i> -value
Season *	-3.663	0.331	1	159.35	16.68	0.0002
Head-capsule width (mean)	5.589	0.801	1	105.52	11.04	0.0012
(a) Each strain included in the model separately						
wExe1 prevalence †	10.305	0.863	1	186.97	7.77	0.0076
wExe2 prevalence †	3.356	0.355	1	100.73	8.08	0.0066
wExe3 prevalence †	0.371	0.645	1	0.32	0.01	0.9056
(b) All 3 strains included in the model together						
wExe1 prevalence †	9.603	0.907	1	151.34	15.84	0.0002
wExe2 prevalence †	4.696	0.537	1	90.32	9.45	0.0036
wExe3 prevalence †	5.923	0.887	1	39.16	4.10	0.0489
(c) Number of strains						
Number of Wolbachia strains	2.160	0.161	1	289.05	60.61	<0.0001

^{* 2} levels: 2007/08 vs 2009/10 and 2010/11; † square-root transformed

The dependent variable in this analysis was the prevalence of virus-dead larvae observed across each of 59 outbreaks; eight observations were excluded due to missing data (see legend to Table S1 for further details). In contrast to Table S1, this version of the model tested the significance of individual *Wolbachia* strains. Final model with all three strains included: intercept = -15.387 ± 1.960; null deviance = 848.60 (50 d.f.); residual deviance = 152.29 (45 d.f.).

Table S3. Determinants of viral load in *S. exempta* across outbreaks.

Term	Coef	s.e.m.	d.f.	Δ deviance	F	<i>P</i> -value
Larval density	-0.0055	0.0024	1	8.00	6.49	0.0138
Outbreak number	-0.0082	0.0393	1	12.97	10.53	0.0021
Wolbachia prevalence *	-1.7481	1.3588	1	6.10	4.96	0.0306
Outbreak no : Wolbachia	0.3642	0.1172	1	11.89	9.66	0.0024
Excluded terms						
Season †			1	1.02	0.36	0.3627
Head-capsule width (mean)			1	0.39	0.41	0.5239
Date ‡			1	11.38	1.27	0.2652

^{*} square-root transformed; † 2 levels: 2007/08 vs 2009/10 and 2010/11; ‡ days since January 1st.

The dependent variable in this analysis was the mean \log_{10} -transformed viral load of apparently healthy larvae observed across each of 59 outbreaks; two observations were excluded due to missing data. Viral load was determined using SpexNPV-specific qPCR. Analysis was by general linear model (GLM with Gaussian error and identity link). Final model: intercept = 3.592 ± 0.522 ; null deviance = 102.92 (56 d.f.); residual deviance = 64.05 (52 d.f.).

Table S4. SpexNPV bioassay mortality analysis.

Term	Coef	s.e.m.	d.f.	Δ deviance (X^2)	<i>P</i> -value
Bioassay *	1.117	0.238	1	24.22	<0.0001
Viral dose †	1.918	0.238	1	133.34	<0.0001
Wolbachia status ‡	1.711	0.239	1	58.81	<0.0001
Excluded terms					
Bioassay : Virus dose			1	1.01	0.3134
Bioassay : Wolbachia status			1	1.37	0.5053
Virus dose : Wolbachia status			1	0.13	0.7210

* 3 separate bioassays, two (bioassays 1 and 3) comparing mortality of wExe1 infected larvae with larvae made Wolbachia-free via tetracycline treatment ($wExe1_{tet}$) and one (bioassay 2) comparing with wExe1-infected larvae with larvae from a line naturally free of Wolbachia; model simplification revealed that bioassays 1 and 3 could be combined without reducing the amount of variation explained. † log_{10} -transformed; ‡ Wolbachia-free or Wolbachia-infected.

The dependent variable in this analysis was the binary indicator of mortality or otherwise of individual 4th instar larvae inoculated with one of a range doses of SpexNPV (0, 100, 500, 1000 and 5000 OB). Analysis was by generalized linear model with binomial errors and logit link function. Parameter estimates in the minimal model are for explanatory terms when fitted last in the model. All other terms were excluded with p > 0.05 when fitted last in the model. Final model: intercept = -7.216 \pm 0.715; null deviance = 717.85 (574 d.f.); residual deviance = 492.24 (574 d.f.).

Table S5. SpexNPV bioassay summary data.

Bioassay	Comparison *	LD ₅₀ (OB/larva) Wol+ †	LD ₅₀ (OB/larva) Wol-‡	Virus yield (OB/larva) Wol+ §	Virus yield (OB/larva) Wol-§
1	wExe1 (line 1) vs wExe1 _{tet} (gen 2)	659	9,370	7.14 x 10 ⁷	9.78 x 10 ⁷
2	wExe1 (line 2) vs Wolbachia-free stocks (gen 2)	196	1,539	7.38 x 10 ⁷	8.72 x 10 ⁷
3	wExe1 (line 1) $vswExe1_{tet} (gen 3)$	1,130	6,702	8.28 x 10 ⁷	10.57 x 10 ⁷

* 3 separate bioassays, two (bioassays 1 and 3) comparing mortality of *wExe1* infected larvae with larvae made *Wolbachia*-free via tetracycline treatment (*wExe1*_{tet}) and one (bioassay 2) comparing with *wExe1*-infected larvae with larvae from a line naturally free of *Wolbachia*; † *Wolbachia*-infected; ‡ *Wolbachia*-free; § mean number of virus occlusion bodies (OB) produced per infected cadaver.

Table S6. SpexNPV bioassay survival analysis.

Term	Coef	s.e.	d.f.	Δ deviance (X^2)	<i>P</i> -value
Bioassay *	0.842	0.165	1	26.01	<0.0001
Viral dose †	1.267	0.157	1	71.67	<0.0001
Wolbachia status ‡	1.222	0.164	1	61.31	<0.0001
Excluded terms					
Bioassay : Virus dose			1	0.02	0.8865
Bioassay : Wolbachia status			1	0.06	0.7973
Virus dose : Wolbachia status			1	0.72	0.3977

* 3 separate bioassays, two (bioassays 1 and 3) comparing mortality of *wExe1* infected larvae with larvae made *Wolbachia*-free via tetracycline treatment (*wExe1*_{tet}) and one (bioassay 2) comparing with *wExe1*-infected larvae with larvae from a line naturally free of *Wolbachia*; model simplification revealed that bioassays 1 and 3 could be combined without reducing the amount of variation explained. † log₁₀-transformed; ‡ *Wolbachia*-free or *Wolbachia*-infected.

The dependent variable in this analysis is the censored survival time from the bioassay experiment. Thus, the response is the time to death where larvae that survival were censored at day 11 post-infection (by which time all survivors had pupated). Analysis was by Cox's proportional hazards model. Parameter estimates in the minimal model are for explanatory terms when fitted last in the model. All other terms were excluded with P > 0.05 when fitted last in the model. Final model: Likelihood ratio test: $X_3^2 = 173$, P < 0.0001, P = 0.293.

Table S7. Determinants of viral loads in bioassay experiments.

Term	Coef	s.e.	d.f.	Δ deviance	F	<i>P</i> -value
Speed of kill *	-4.769	0.5684	1	71.41	16.16	<0.0001
Wolbachia status †	-0.193	0.0319	1	35.79	8.10	0.0050
Excluded terms						
Virus dose ‡			1	1.73	0.39	0.5330
Bioassay			2	10.61	1.19	0.3068
Wolbachia status: Speed			1	5.03	1.14	0.2867

^{*} speed of kill = 1 / number of days taken to die post infection; † Wolbachia-free or Wolbachia-95 96

The dependent variable in this analysis was the count of virus occlusion bodies (OB) from SpexNPV-killed larvae in the bioassays. Analysis was by generalised linear model (GLM with Poisson errors and log link). To account for over-dispersion, an empirical dispersion parameter and F-tests were used. Final model: intercept = 4.032 ± 0.088 ; null deviance = 858.84 (179 d.f.); residual deviance = 745.89 (177 d.f.). Using these lab-generated data for virus yield, we can calculate a lab-based proxy for the basic reproductive rate of the virus (R_0) , based on the production of occluded virus only. This was done by multiplying the mortality rate of the larvae for a given dose of virus by the average yield of virus occlusion bodies for larvae infected with that dose across each of the three bioassays. The comparison of R_0 is made simply by dividing the estimate for Wol+ larvae by that for Wol- larvae for each dose. Averaged across the three bioassays, R_0 ratios (Wol+/Wol-) for the four viral doses are as follows: 100 OB = 2.93 (range = 1.16 - 4.10), 500 OB = 2.96 (2.00 - 4.40), 1000 OB = 1.60 (1.05 - 1.87), 5000 OB = 1.27(Bioassay 2 only).

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infected; ‡ log₁₀-transformed.

Table S8. Microbial species infecting *Spodoptera exempta* in outbreak populations.

A list of the microbial species identified in African armyworm samples using PCR-

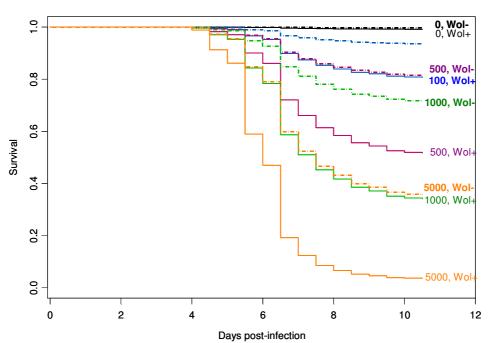
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Isolate	BLASTN	Bacteria genus	Nucleotide	Habitat isolated
no.	match		% identity	
1	FJ390735.1	Uncultured bacterium	91%	soil
2	GQ489023.1	Enterococcus faecium	100%	environment
3	GQ453520.1	Enterococcus faecium	100%	environment
4	GQ299825	Uncultured bacterium	100%	environment
5	GQ383919.1	Bacillus sp.	100%	environment
6	FJ854700.1	Uncultured beta proteobacterium	100%	environment
7	GQ426315.1	Pantoea sp.	98%	rice plant
8	GQ383917.1	Paenibacillus sp.	98%	soil
9	AB522100.1	Kineococcus radiotolerans	100%	soil
10	GQ495647.1	Curtobacterium sp.	100%	soil

Figure S1. Survival of *S. exempta* larvae from *Wolbachia*-infected (Wol+) and

Wolbachia-free (Wol-) lines following inoculation with SpexNPV.

Expected survival (specified covariates)



Solid lines are the survival curves for *Wolbachia*-infected lines; dashed lines are for *Wolbachia*-free lines. Different colours reflect different infecting doses (numbers next to lines).